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Cary - Blair Medium Base (Transport Medium w/o Charcoal)

Recommended for collection and shipment of clinical specimens.

REF: BS.1/CMP01.100.0100	100 Gram	REF: BS.1/CMP01.250.0250	250 Gram
REF: BS.1/CMP01.500.0500	500 Gram		

CLINICAL SIGNIFICANCE

Transport Medium is a non-nutritive, chemically defined, semisolid, buffered medium. The sole purpose of this medium is to maintain the viability of organisms during the time from collection to examination of the specimen. Transport Medium should be essentially non-nutritive so that the test organisms do not increase in numbers during transport. Transport media were originally formulated by Stuart et al (1) for carrying gonococcal specimens to the laboratory. Cary and Blair devised a new medium containing fewer nutrients, low oxidation-reduction potential and a high pH (2). Cary-Blair Medium w/o Charcoal is used for collection and transport of clinical specimens. It is also recommended by APHA (3) and various authors for transport of specimens (4,5,6). Since this transport media has a high pH, viability of Vibrio cultures can be maintained for a longer duration (7). This medium also facilitates the recovery of Salmonella and Shigella species (4).

METHOD PRINCIPLE

Cary-Blair Medium Base is prepared with minimal nutrients to facilitate survival of organisms without multiplication. Sodium thioglycollate provides a low oxidation-reduction potential. Alkaline pH of the medium minimizes bacterial destruction due to the formation of acid. Disodium hydrogen phosphate buffers the medium whereas sodium chloride maintains the osmotic equilibrium. For collection of the specimen, use sterile cotton tipped swabs on wooden sticks. Push the swabs down to one third of the medium depth and cut the stick so that when the cap is screwed down, the swab is forced to the bottom of the medium. Tighten the cap firmly on the bottle. The specimen will be preserved and the viability of the organisms will be also maintained during transport, but over the time it will diminish. Therefore direct inoculation of the specimen is advised. Some growth of accompanying contaminants may also occur during longer period of transit. The specimen should be inoculated into a proper medium as soon as possible.

MEDIA COMPOSITION

Item	Formula per liter of medium
- Disodium hydrogen phosphate - Sodium thioglycollate - Sodium chloride - Agar	1.100 gm 1.500 gm 5.000 gm 5.000 gm

Final pH 8.4 ± 0.2 at 25 °C

PRECAUTIONS AND WARNINGS

Media to be handled by entitled and professionally educated person. Do not ingest or inhale.

Good Laboratories practices using appropriate precautions should be followed in:

- Wearing personnel protective equipment (overall, gloves, glasses,).
- Do not pipette by mouth.
- In case of contact with eyes or skin; rinse immediately with plentyof soap and water. In case of severe injuries; seek medical adviceimmediately.
- Respect country requirement for waste disposal.

S56: dispose of this material and its container at hazardous or special waste collection point.

S57: use appropriate container to avoid environmental contamination.

S61: avoid release in environment.

For further information, refer to the Cary-Blair Medium Base material safety datasheet.

STORAGE AND STABILITY

BioScien Cary-Blair Medium Base should be stored between 10-30°C in a firmlyclosed container and the prepared medium at 2-8°C. Use before expiry date on the label. On opening, product should be properly stored dry, after tightly capping the bottle in order to avoid lump development due to the hygroscopic nature of the product. Improper storage of the product may lead to lump formation. Store in a dry ventilated area protected from extremes of temperature andsources of ignition. Seal the container tightly after use. Product performance is best if used within stated expiry period.

PREPARATION

Suspend 12.6 grams in 991 mlpurified/distilled water. Heat to boiling to dissolve the medium completely. Cool to 45-50°C and aseptically add 9 ml of 1% aqueous calcium chloride solution. Adjust pH to 8.4, if necessary. Distribute in 7 ml amounts in screw capped tubes. Steam for 15 minutes. Cool and tighten the caps.

Deterioration

The color of ***BioScien*** Cary-Blair Medium Base is Cream to yellow homogeneous free flowing powder. prepared medium is Light amber coloured, slightly opalescent solution in tubes. If there are any physical changes for powder or signs of deterioration (shrinking, cracking, or discoloration), and contaminations for hydrated media, discard the medium.

EQUIPMENT REQUIRED NOT PROVIDED

- Sterile cups
- Sterile petri-dishes
- Incubator

SPECIMEN

Clinical samples : faeces, urine, Nasopharyngeal swabs etc

PERFORMANCE CHARACTERISTICS

Cultural characteristics observed after an incubation at 35-37°C for 18-24 hours, when subcultured on Tryptone Soya Agar.

Organisms	Growth
Klebsiella aerogenes ATCC 13048	Good-luxuriant
Escherichia coli ATCC 25922	Good-luxuriant
Neisseria meningitidis ATCC 13090	Good-luxuriant
Salmonella Typhimurium ATCC 14028	Good-luxuriant
Vibrio parahaemolyticus ATCC 17802	Good-luxuriant

SYMBOLS IN PRODUCT LABELLING	
LOT Batch Code/Lot number	Caution
REF Catalogue Number	Do not use if package is damaged
Temperature Limitation	Consult Instruction for use
Expiration Date	
Manufactured by	

QUALITY CONTROL

To ensure adequate quality control, it is recommended that positive and negative control included in each run. If control values are found outside the defined range, check the system performance. If control still out of range please contact ***BioScien*** technical support.

REFERENCES

1.Stuart, Toshach and Pastula, 1954, Can. J. Public Health, 45:73.
2. Cary and Blair, 1964, J. Bacteriol., 88:96.
3. Salfinger Y., and Tortorello M.L., 2015, Compendium of Methods for the Microbiological Examination of Foods, 5th Ed., American Public Health Association, Washington, D.C.
4. Cary, Mathew, Fusillo and Harkins, 1965, Am. J. Clin. Pathol., 43:294
5. Gaines et al, 1965, Am. J. Trop. Med. Hyg., 14:136.
6.Morris and Heck, 1978, J. Clin. Microbiol., 8:616.
7. Murray P. R., Baron E. J., Tenover F. C., Pfaller M. A., Yolken R.H., (Eds.), 8th Ed., 2003, Manual of Clinical Microbiology, ASM, Washington, D.C.
8.Isenberg, H.D. Clinical Microbiology Procedures Handbook 2nd Edition.
9.Jorgensen, J.H., Pfaller, M.A., Carroll, K.C., Funke, G., Landry, M.L., Richter, S.S and Warnock., D.W. (2015) Manual of Clinical Microbiology, 11th Edition. Vol. 1.